

Impact of Temperature on Heparin and Protein Interactions

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Abstract

Heparin has many important biological activities, associated with a diverse set of interactions with biologically functional proteins. The binding mechanisms and biological significance of heparin-protein interactions have attracted wide attention. However, the temperature sensitivity of heparin-protein interaction is relatively unstudied. The impact of temperature on the binding of heparin to three representative heparin-binding proteins, antithrombin III (AT III), fibroblast growth factor-1 (FGF1) and fibroblast growth factor-2 (FGF2) are evaluated. The affinity and kinetics of these interactions were measured at 10°C, 25°C and 30°C. The association rate, dissociation rate, binding affinity and binding mass were compared at different temperatures. In the two state binding process between AT III and heparin, temperature played a negligible role on ATIII binding to heparin (1st state reaction), but demonstrated a role in the conformational change process (2nd state reaction). In the case of FGF1 and FGF2, the kinetics and affinity, while distinctly different at the temperatures studied, were still within the same order of magnitude. Based these results, we conclude that in many cases it is possible to perform surface plasmon resonance measurements of heparin-protein interaction at different temperatures, especially at reduced (ambient or lower) temperatures, and obtain comparable binding data.

Keywords: Heparin; Protein; Interaction; Temperature; Surface plasmon resonance

Abbreviations: GAG: Glycosaminoglycan; SPR: Surface Plasmon Resonance; HS: Heparan Sulfate; ECM: Extracellular Matrix; FGFs: Fibroblast Growth Factors; FGFRs: Fibroblast Growth Factor Receptors; AT III: Antithrombin III; FGF1: Fibroblast Growth Factor-1; FGF2: Fibroblast Growth Factor-2; SA: Streptavidin; FC: flow-Cell; RU: Resonance Unit; k_a : Association Rates; k_d : Dissociation Rates; K_D : Dissociation Constant. SD: Standard Deviations.

Introduction

Heparan Sulfate (HS) and heparin are the most structurally complex members of the Glycosaminoglycan (GAG) family. These anionic, polydisperse, linear polysaccharides are highly sulfated and are O-glycosidically linked to a core protein to comprise HS and heparin proteoglycans (PGs). HS, found on the external cell membrane and also in the Extracellular Matrix (ECM), can interact with many different protein ligands [1-3]. Many physiological and pathophysiological processes can be modulated through heparin/HS-protein interactions including: blood coagulation, cell growth and differentiation, host defense, viral infection, lipid transport and metabolism, cell-to-cell and cell-to-matrix signaling, inflammation, Alzheimer's disease and cancer [1]. HS-protein interactions have been demonstrated to critically modulate the biological activity of the proteins through a variety of mechanisms, including localization of proteins in the ECM, regulation of enzymatic activities, ligands binding to receptors, and protein protection against proteolysis [2]. Heparin/HS-PGs are also essential cofactors required for the interaction of fibroblast growth factors (FGFs) with their protein-based receptors (FGFRs) [3,4].

Biomolecular interactions are governed by weak forces including hydrogen bonds, van der Waals forces, ionic interactions, and hydrophobic interactions. Biological systems are usually restricted to a narrow range of physical conditions, since biomacromolecules are functionally active only within a narrow range of environmental conditions, such as temperature, ionic strength, and pH. Extremes of these conditions can overcome the weak forces essential to maintaining the intricate structure of macromolecules [5]. The effect of temperature on heparin-protein interactions is, however, rarely discussed, although

heparin can occupy new vibrational modes as the temperature increases [6]. In traditional binding assays, the affinity measurements of protein interactions are typically performed at room temperature or lower, with little or no reflection of the potential impact of the temperature on these interactions. In the present study, the effect of temperature on the binding of heparin and three representative bioactive proteins was evaluated, and the affinity and kinetics of the interactions were analyzed at these selected temperatures. Changes in the behavior of the interactions, particularly in the association and dissociation rates were observed.

Materials and Methods

Materials

Porcine mucosal heparin (~16 kDa) was from Celsus Laboratories, (Cincinnati, OH). Human antithrombin III (AT III) was from Haematologic Technologies (Essex Junction, Vermont). FGF1 and FGF2 were generously provided by Amgen (Thousand Oaks, CA). GE Healthcare Bio-Sciences AB (Uppsala, Sweden) was the source of the SA sensor chips. A BIAcore 3000, running under BIAcore 3000 control and Version 4.0.1 BIAevaluation software, was used for SPR measurements.

Heparin biochip preparation

Biotin-heparin conjugate was synthesized through the reaction

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of long-chain sulfo-N-hydroxysuccinimide biotin (Thermo Fisher, Rockford, IL) with the unsubstituted amino groups of the polysaccharide chain's glucosamine residues based on published methods [7-9]. The biotin-heparin conjugate was then bound to a Streptavidin (SA) chip. A 20 μ L solution of the biotin-heparin conjugate in HBS-EP running buffer (0.1 mg/mL) was injected over SA chip flow cell 2 at 10 μ L/min. Immobilization of heparin was indicated through a \sim 250 resonance unit (RU) increase. Control Flow Cell (FC₁) was prepared through a 1 min injection of saturated biotin.

Measuring heparin-protein interaction using SPR at different temperatures

Protein samples were made up in HBS-EP buffer (0.01 M 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid, 0.15 M sodium chloride, 3 mM ethylene diaminetetraacetic acid, 0.005% surfactant P20, pH 7.4). Protein samples at different concentrations were injected to the heparin chip at 30 μ L/min. The HBS-EP buffer was flowed over the sensor surface to facilitate dissociation at the end of the sample injection. After dissociation over a 3 min, the surface of the sensor was completely regenerated with 30 μ L of 2 M NaCl. Sensorgrams measured response as a function of time and were recorded at 10, 25 and 30 $^{\circ}$ C.

Results and Discussion

Kinetics measurement of protein-heparin interactions under different temperatures

The kinetics and affinity of the interactions between heparin and three biologically important heparin-binding proteins (HBP) were measured by SPR under different temperatures. AT III is a serine protease inhibitor that inactivates various activated coagulation serine proteases, including factors IXa, Xa, TF-VIIa complex, and thrombin (factor IIa) [10,11]. The anticoagulant activity of heparin is primarily mediated through its binding and regulation of AT III [12-14], making the interaction between heparin and AT III a crucial step in the anticoagulation process. FGF1 and FGF2, also known as acidic and basic fibroblast growth factor, respectively, belong to the FGF family having 23 members and play an important role in the cell proliferation, differentiation, morphogenesis and angiogenesis, with heparin binding events highly involved [15-18].

Evaluation of heparin interaction with AT III, FGF1 and FGF2 was conducted under 10 $^{\circ}$ C, 25 $^{\circ}$ C and 30 $^{\circ}$ C, and the association rates (k_a), dissociation rates (k_d) and dissociation constant (K_D) were calculated. SPR sensorgrams of heparin-protein interactions at different temperatures were shown in Figures 1-3. Kinetics and affinity data are summarized in Table 1.

For AT III binding, the sensorgrams of heparin-ATIII interaction fit very well to a two-state binding model (Figure 1), indicating that ATIII undergoes a conformational change after it binding to heparin. The conformational change in ATIII on heparin binding is well established and is important in ATIII-mediated inhibition of many serine proteases and anticoagulant activity of heparin [19]. It appears that temperature has minimal effects on the ATIII binding to heparin (1st state reaction) since the measured k_{a1} and k_{d1} values were not obviously different at 10, 25, and 30 $^{\circ}$ C. It is interesting to see that the conformational change process (2nd state reaction) was impacted by temperature apparently, since the k_{d2} decreased with the increased temperature with $k_{d2} = 4.78 \times 10^{-4} (\pm 1.1 \times 10^{-6})$ (1/S) at 10 $^{\circ}$ C, $k_{d2} = 2.34 \times 10^{-4} (\pm 6.7 \times 10^{-7})$ (1/S) at 25 $^{\circ}$ C, and $k_{d2} = 7.9 \times 10^{-7} (\pm 5.3 \times 10^{-9})$ (1/S) at 30 $^{\circ}$ C. These results suggest that a higher temperature favors conformational change and

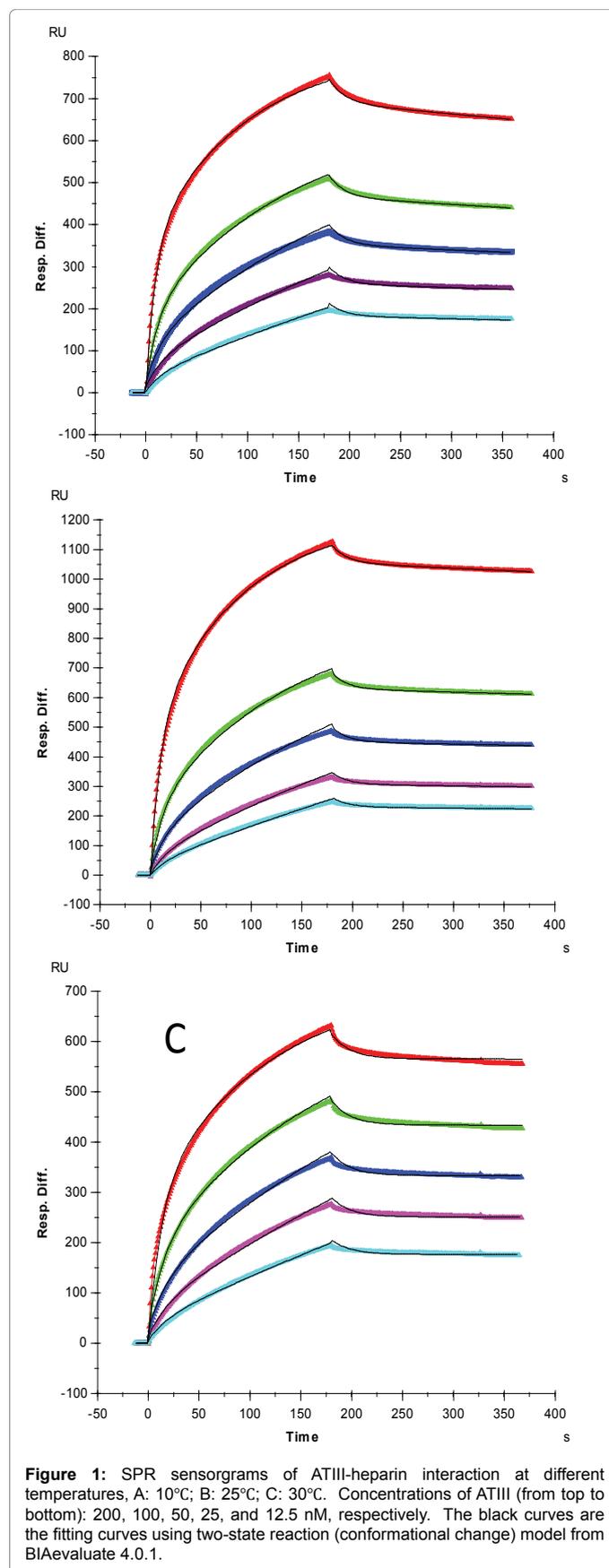


Figure 1: SPR sensorgrams of ATIII-heparin interaction at different temperatures, A: 10 $^{\circ}$ C; B: 25 $^{\circ}$ C; C: 30 $^{\circ}$ C. Concentrations of ATIII (from top to bottom): 200, 100, 50, 25, and 12.5 nM, respectively. The black curves are the fitting curves using two-state reaction (conformational change) model from BIAevaluate 4.0.1.

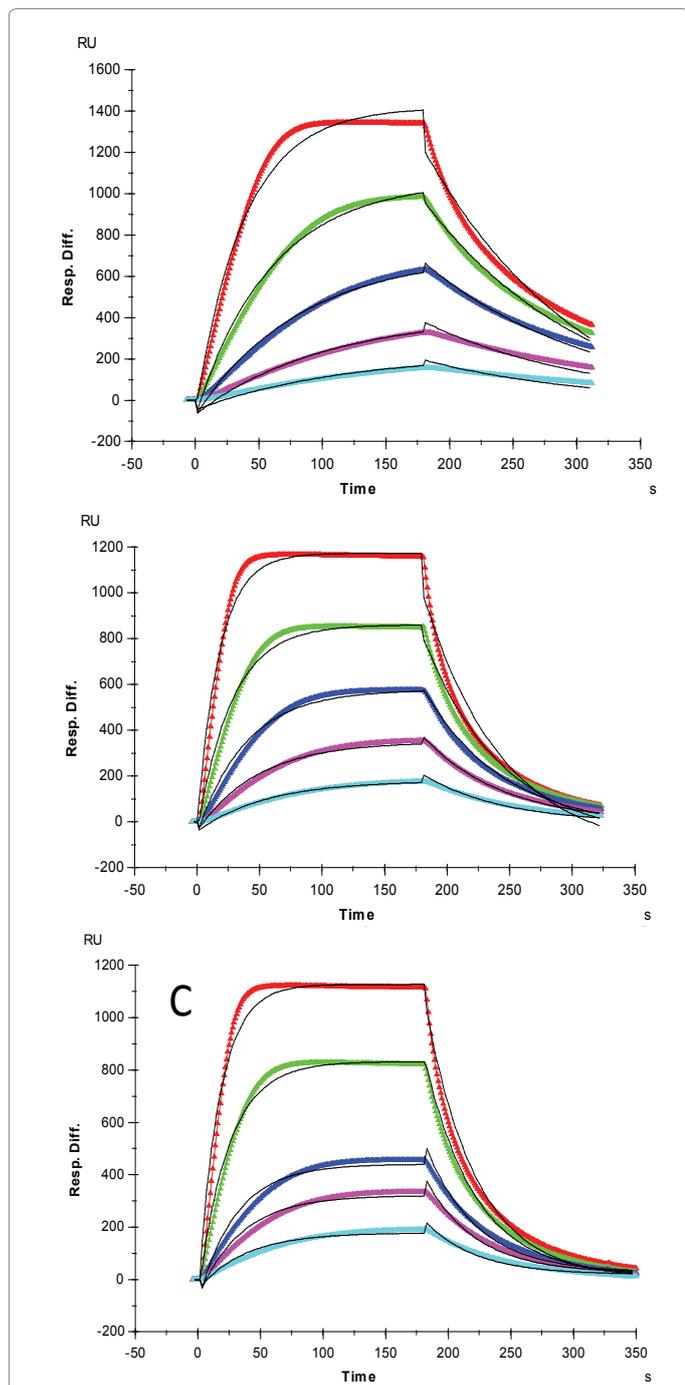


Figure 2: SPR sensorgrams of FGF1-heparin interaction at different temperatures, A: 10°C; B: 25°C; C: 30°C. Concentrations of FGF1 (from top to bottom): 200, 100, 50, 25, and 12.5 nM, respectively. The black curves are the fitting curves using 1:1 Langmuir binding model from BIAevaluate 4.0.1.

gives more stable ATIII-heparin complex. It was found that heparin produces a decrease in the β -structure of ATIII and a compensatory increase in random coil [20], which determines that the conformational change process is an entropy increased ($\Delta S > 0$) event. In such case, the increase of temperature would induce a lower Gibbs free energy (ΔG), giving higher tendency to form the conformation changed ATIII-heparin complex, which was testified by the significantly lower value of k_{d2} we got at 30°C. From another point of view, since hydrophobic

interaction, which is the main force for protein folding, was found to be weaker at lower temperature [21], the stability of ATIII-heparin complex might be reduced at lower temperature, corresponding to the higher k_{d2} at 10°C and 25°C.

For FGF1 and FGF2, 1:1 Langmuir binding models were used for fitting. The lower reaction rates were expected at lower temperature since higher temperatures supply more energy, potentially leading to an accelerated interaction between heparin and proteins. An increase of both k_a and k_d was observed for FGF1 and FGF2 with temperature

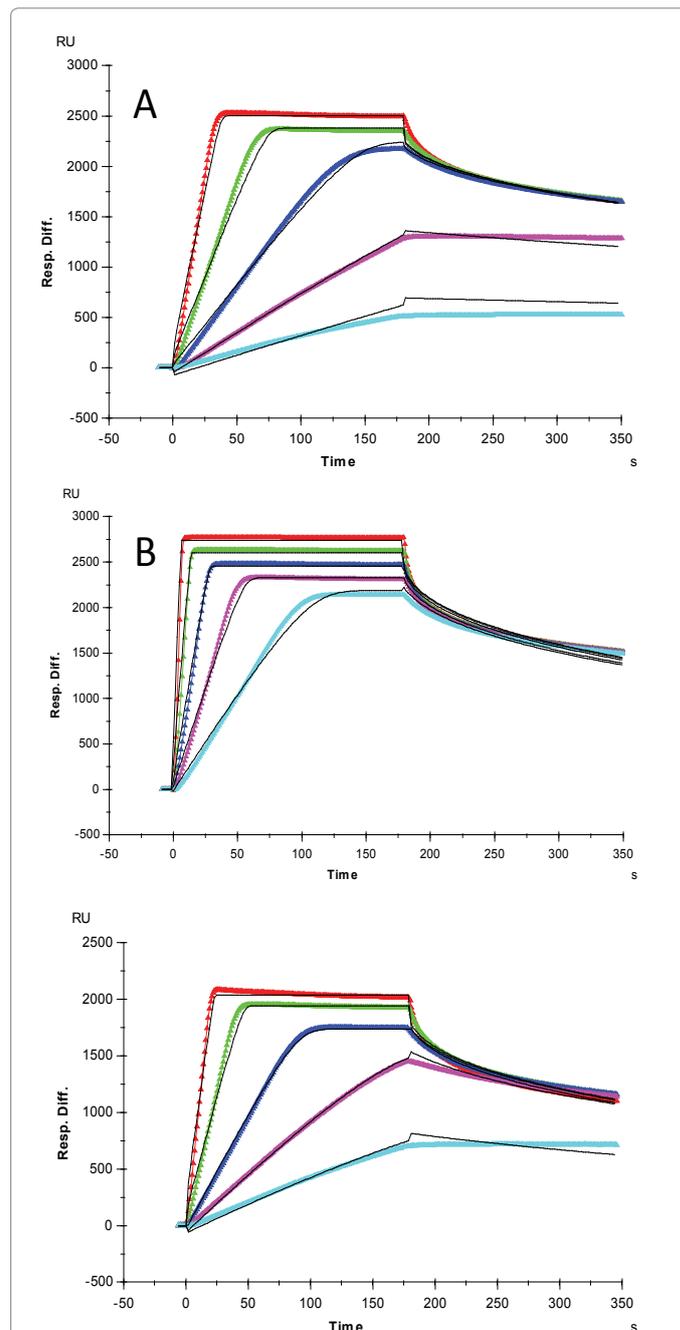


Figure 3: SPR sensorgrams of FGF2-heparin interaction at different temperatures, A: 10°C; B: 25°C; C: 30°C. Concentrations of FGF2 (from top to bottom): 200, 100, 50, 25, and 12.5 nM, respectively. The black curves are the fitting curves using 1:1 Langmuir binding model from BIAevaluate 4.0.1.

Protein	Temperature	k_a (1/MS)	k_d (1/S)	K_D (M)
ATIII	10°C	$k_{a1} = 2.63 \times 10^5 (\pm 437)$ (1/MS)	$k_{d1} = 0.047 (\pm 1.0 \times 10^{-4})$ (1/S)	
		$k_{a2} = 0.021 (\pm 3.7 \times 10^{-5})$ (1/S)	$k_{d2} = 4.78 \times 10^{-4} (\pm 1.1 \times 10^{-6})$ (1/S)	
	25°C	$k_{a1} = 2.4 \times 10^5 (\pm 433)$ (1/MS)	$k_{d1} = 0.043 (\pm 1.1 \times 10^{-4})$ (1/S)	
		$k_{a2} = 0.024 (\pm 5.1 \times 10^{-5})$ (1/S)	$k_{d2} = 2.34 \times 10^{-4} (\pm 6.7 \times 10^{-7})$ (1/S)	
	30°C	$k_{a1} = 1.9 \times 10^5 (\pm 886)$ (1/MS)	$k_{d1} = 0.035 (\pm 2.4 \times 10^{-4})$ (1/S)	
		$k_{a2} = 0.019 (\pm 8.1 \times 10^{-5})$ (1/S)	$k_{d2} = 7.9 \times 10^{-7} (\pm 5.3 \times 10^{-9})$ (1/S)	
FGF1	10°C	9.1×10^4 ($\pm 6.6 \times 10^3$)	7.6×10^{-3} ($\pm 4.7 \times 10^{-5}$)	8.3×10^{-8}
		25°C	2.0×10^5 ($\pm 6.6 \times 10^3$)	0.017 ($\pm 4.7 \times 10^{-5}$)
	30°C		2.0×10^5 ($\pm 6.6 \times 10^3$)	0.018 ($\pm 4.7 \times 10^{-5}$)
		10°C	4.6×10^7 ($\pm 6.6 \times 10^3$)	0.102 ($\pm 4.7 \times 10^{-5}$)
	25°C		3.8×10^8 ($\pm 6.6 \times 10^3$)	0.406 ($\pm 4.7 \times 10^{-5}$)
		30°C	9.2×10^7 ($\pm 6.6 \times 10^3$)	0.296 ($\pm 4.7 \times 10^{-5}$)

Table 1: Summary of kinetic data of heparin-protein interaction at different temperatures*.

elevated from 10°C to 25°C or 30°C. For FGF1, k_a was 9.1×10^4 (1/MS) at 10°C, and 2.0×10^5 (1/MS) at 25°C and 30°C, in the meantime k_d increased from 7.6×10^{-3} (1/S) at 10°C to 0.017 (1/S) and 0.018 (1/S) at 25°C and 30°C, respectively. As k_a and k_d both increased by temperature, the affinity K_D which was calculated by equation ($K_D = k_d / k_a$) became inestimable. For FGF1, the same value of K_D (8.3×10^{-8} M) was obtained at 10°C and 25°C, implying a common binding affinity at different temperatures. This represents a good case to demonstrate that despite of the diverse kinetics between heparin and protein, binding affinity might be similar at different temperatures since the change of k_a was compensated by the change of k_d , when they were in the same direction. In contrast, it is interestingly noted that at 30 °C, K_D of heparin-FGF1 binding was calculated to be 1.1×10^{-7} M, indicating a lower binding affinity than those at lower temperatures. This could be interpreted as the binding affinity was dominated by the faster dissociation rate at 30°C.

FGF2 had higher affinity with heparin and distinct binding kinetics, showing a “fast association and fast dissociation” mode (Table 1, Figures 2 and 3). The k_a of FGF2 was approximately three orders of magnitudes larger than the k_a of FGF1, while k_d was only about two orders of magnitudes lower, resulting in a smaller K_D (higher affinity) for FGF2. Similarly, an increase of both k_a and k_d was observed with temperature elevated from 10°C to higher. The k_a for FGF2 was 4.6×10^7 (1/MS) at 10°C, 3.8×10^8 (1/MS) at 25°C and 9.2×10^7 (1/MS) at 30°C, in the same time k_d increased from 0.102 (1/S) at 10°C to 0.406 (1/S) and 0.296 (1/S) at 25°C and 30°C, respectively. However, FGF2 showed a significant decline of both k_a and k_d when temperature was increased from 25°C to 30°C, leading to the K_D (3.2×10^{-9} M) at 30°C larger (lower affinity) than that at 25°C (1.1×10^{-9} M) and even 10°C (2.2×10^{-9} M). The dramatic changes of binding kinetics between FGF2

and heparin might be caused by the lower bioactivity and reduced stability of FGF2 at 30°C.

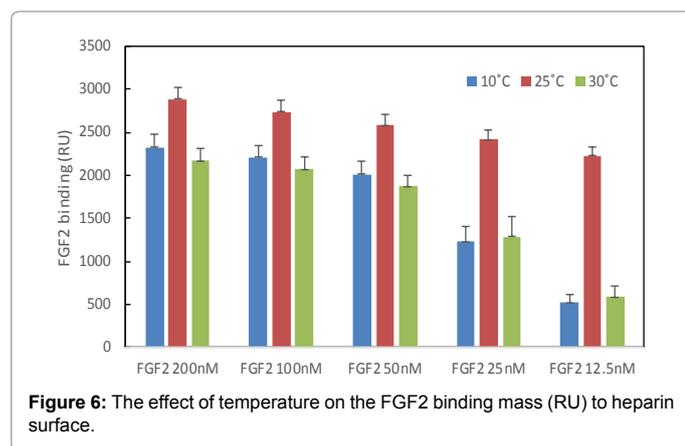
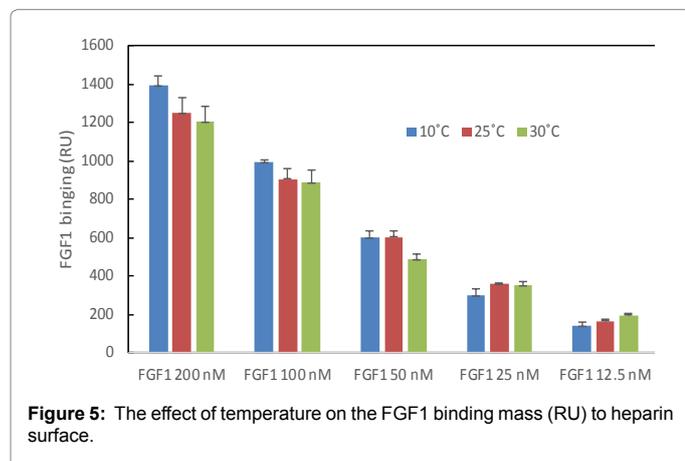
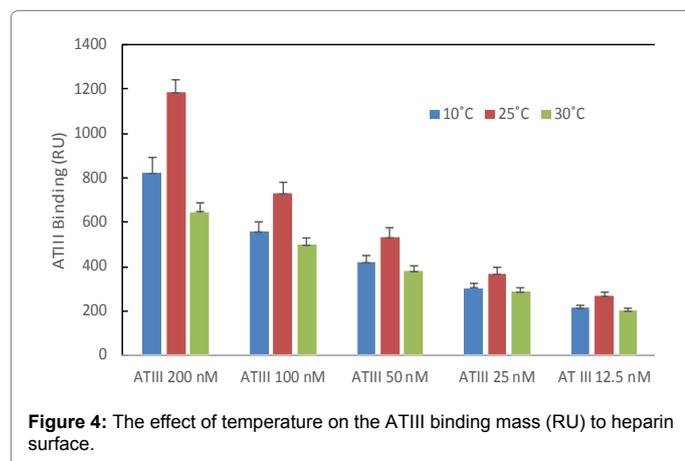
It is important to note that, for each of the protein tested, even though the kinetics and affinity were different at different temperatures, they were still in the same order of magnitude. Based on these results, we conclude that it is possible to conduct measurements of heparin-protein interaction at different temperatures by SPR, especially at low (ambient or lower) temperature and that influence of reduced temperature on the interaction behavior is likely to be low (less than one order of magnitude).

Effect of temperature on protein binding mass to heparin surface

The binding masses (RU) of three proteins to heparin surface were recorded at different protein concentrations and different temperatures. The binding mass corresponds to the maximum RU response achieved when certain amount of protein samples is flowed, for a fixed period of time, across the surface of a heparin-immobilized chip. Surprisingly, AT III showed highest binding mass to the chip at 25°C when compared to that obtained at lower (10°C) and higher (30°C) temperatures (Figure 4). Thus, one should carefully consider binding temperature when binding assays are performed under similar physicochemical conditions. As shown in Figure 5, the influence of temperature on binding mass of FGF1 to heparin surface was not significant. While FGF2 was observed to have notably largest RU at 25°C as shown in Figure 6, which was in consistent with the smallest K_D (highest affinity) measured at ambient temperature.

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References

1. Capila I, Linhardt RJ (2002) Heparin-protein interactions. *Angew Chem Int Ed Engl* 41: 391-412.
2. Esko JD, Selleck SB (2002) Order out of chaos: Assembly of ligand binding sites in heparan sulfate. *Annu Rev Biochem* 71: 435-471.
3. Powell AK, Yates EA, Fernig DG, Turnbull JE (2004) Interactions of heparin/heparan sulfate with proteins: Appraisal of structural factors and experimental approaches. *Glycobiology* 14: 17R-30R.
4. Faham S, Hileman RE, Fromm JR, Linhardt RJ, Rees DC (1996) Heparin Structure and Interactions with basic fibroblast growth factor. *Science* 271: 1116-1120.
5. Garrett RH, Grisham CM (2013) *Biochemistry In: Brooks/Cole, Cengage Learning (5th eds) Mary Finch, USA.*
6. Meneghetti MC, Hughes AJ, Rudd TR, Nader HB, Powell AK, et al. (2015) Heparan sulfate and heparin interactions with proteins. *J Royal Soc Interface* 12: 0589.
7. Hernaiz M, Liu J, Rosenberg RD, Linhardt RJ (2000) Enzymatic modification of heparan sulfate on a biochip promotes its interaction with antithrombin III. *Biochem Biophys Res Commun* 276: 292-297.
8. Munoz E, Xu D, Kemp M, Zhang F, Liu J, et al. (2006) Affinity, kinetic, and structural study of the interaction of 3-O-sulfotransferase isoform 1 with heparan sulfate. *Biochemistry* 45: 5122-5128.
9. Zhang F, Liang X, Pu D, George KI, Holland PJ, et al. (2013) Biophysical characterization of glycosaminoglycan-IL-7 interactions using SPR. *Biochimie* 94: 242-249.
10. Bjork I (1989) Molecular mechanisms of the accelerating effect of heparin on the reactions between anti-thrombin and clotting proteinases. *Heparin: Chemical and biological properties, clinical applications* 229.
11. Gray E, Hogwood J, Mulloy B (2012) The anticoagulant and antithrombotic mechanisms of heparin. In: *Heparin-A century of progress*, Springer, Berlin, Heidelberg pp 43-61.
12. Rosenberg RD, Damus PS (1973) The purification and mechanism of action of human antithrombin-heparin cofactor. *J Biol Chem* 248: 6490-6505.
13. Linhardt RJ (1991) Heparin: An important drug enters its seventh decade. *Chem Ind* 2: 45-50.
14. Atha DH, Lormeau JC, Petitou M, Rosenberg RD, Choay J (1985) Contribution of monosaccharide residues in heparin binding to antithrombin III. *Biochemistry* 24: 6723-6729.
15. Ornitz DM (2000) FGFs, heparan sulfate and FGFRs: complex interactions essential for development. *Bioessays* 22: 108-112.
16. Thompson LD, Pantoliano MW, Springer BA (1994) Energetic characterization of the basic fibroblast growth factor-heparin interaction: Identification of the heparin binding domain. *Biochemistry* 33: 3831-3840.
17. Mach H, Volkin DB, Burke CJ, Middaugh CR, Linhardt RJ, et al. (1993) Nature of the interaction of heparin with acidic fibroblast growth factor. *Biochemistry* 32: 5480-5489.
18. Kan M, Wang F, Xu J, Crabb JW, Hou J, et al. (1993) An essential heparin-binding domain in the fibroblast growth factor receptor kinase. *Science* 259: 1918-1921.
19. Linhardt RJ (2003) Claude S Hudson Award address in carbohydrate chemistry. Heparin: Structure and activity. *J Med Chem* 46: 2551-2564.
20. Villanueva GB, Danishefsky I (1977) Evidence for a heparin-induced conformational change on antithrombin III. *Biochem Biophys Res Commun* 74: 803-809.
21. Van Dijk E, Hoogeveen A, Abeln S (2015) The hydrophobic temperature dependence of amino acids directly calculated from protein structures. *PLoS Comput Biol* 11: e1004277.